

PAA









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Microbial and phenyl acid dynamics during the startup phase of anaerobic straw degradation in mesoand thermophilic batch reactors



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Introduction Organic waste materials coming from municipalities or agricultural facilities are valuable substrates for biogas production. One obstacle of using (pretreated) organic wastes is the increased entry of aromatic compounds like lignocellulose or secondary plant metabolites. Aromatic compounds can cause poor biogas production due to intracellular malfunctions and biochemical restrictions¹. However, the effects on AD communities are numerous and depend on several - not fully understood - microbiological and biochemical factors. Previous studies concluded that phenylacetate (PAA) and phenylpropionate (PPA) can be important and early detectable intermediates during AD of aromatic compounds. The objectives of this study were to i) initiate phenyl acid formation during the start-up phase of anaerobic straw degradation, ii) monitor microbial communities during high concentrations of straw from grain and iii) detect biomarkers for high phenyl acid concentrations.

Material and Methods

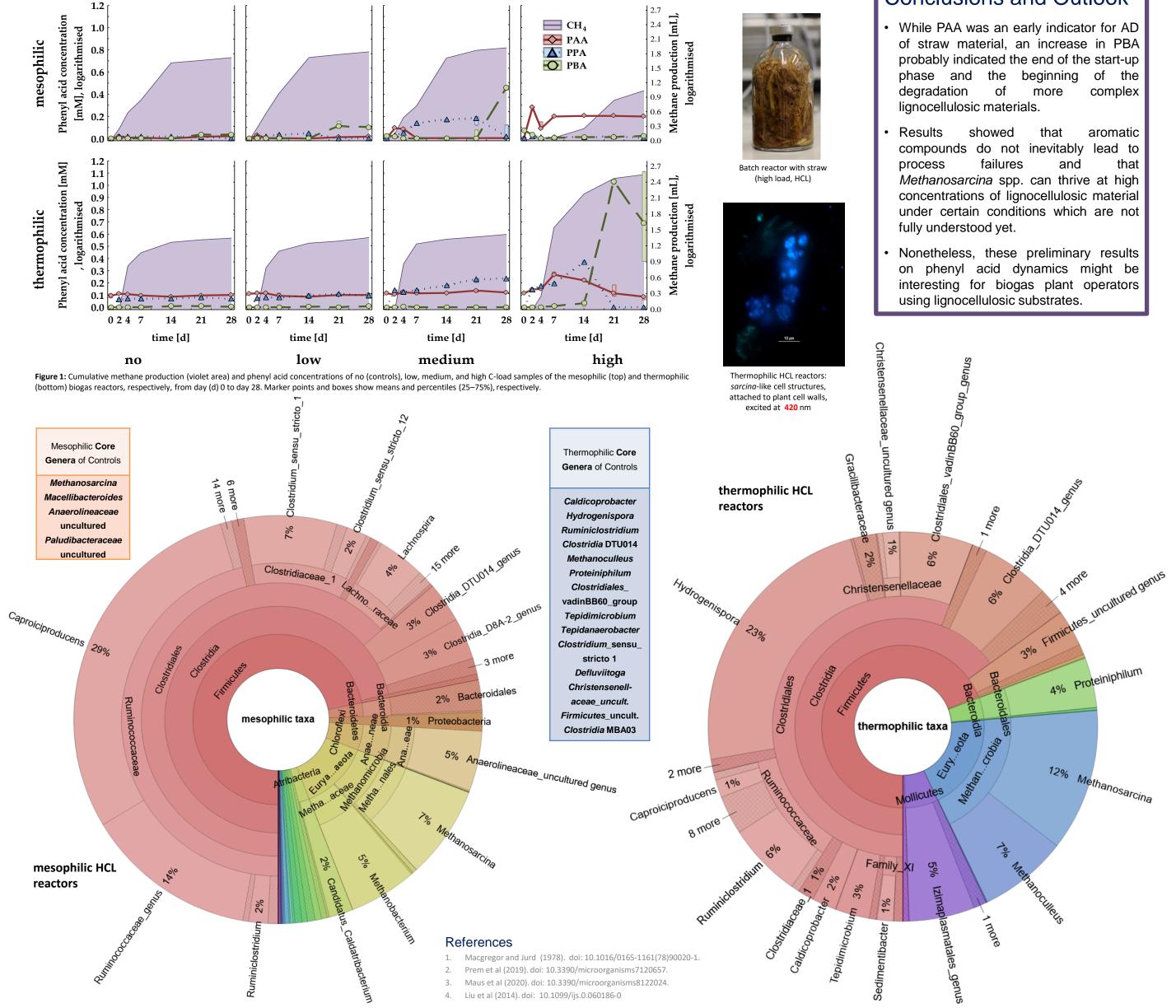
thermophilic batch reactors were set up with different loadings (low: LCL, medium: MCL, high: HCL) of straw from grain as

Meso- and

described in Prem et al.². Gas as well as phenyl acid concentrations were measured chromatographically and the microbial community structure was analysed via high throughput amplicon sequencing of the 16S rRNA.

Results On day 28, mesophilic MCL and thermophilic HCL samples produced significantly more methane than the control and LCL samples, respectively (Fig. 1). A cascade-like pattern – from PAA to PPA to phenylbutyrate (PBA) - was observed for meso- and especially for thermophilic samples. Mesophilic reactors: The highest phenyl acid concentrations were detected in MCL and HCL samples with $123 \pm 3 \text{ mg } L^1$ PAA in HCL samples on day 2, and 73 \pm 3 mg L¹ PPA on day 14 and 307 \pm 24 mg L¹ PBA in MCL samples on day 28. Thermophilic reactors: The highest phenyl acid concentrations were observed in HCL straw samples with 119 \pm 9 mg L¹ PAA on day 7, 202 \pm 8 mg L^1 PPA on day 14, and 1593 ± 80 mg L^1 PBA on day 21.

Methanosarcina – despite its previously described susceptibility to aromatic compounds - was the prevailing methanogenic genus in all mesophilic samples and in thermophilic HCL samples. Methanoculleus spp. was present in all thermophilic samples and was dominating in control, LCL and MCL reactors; the genus might have cooperated with syntrophic acetate oxidisers like Syntrophaceticus spp., Tepidanaerobacter spp. or Clostridia DTU014 genus. Genera like Petrimonas (mesophilic, MCL) and Hydrogenispora (thermophilic, HCL) – representative species are fermentative and produce acetate and hydrogen^{3,4} - were significant biomarkers when methane and phenyl acid production were high.



	C-load	LEfSe Biomarkers
	(Class)	(LDA Score ≥ 4.0)
Mesophilic	control	<i>Anaerolineaceae</i> uncult. genus <i>Clostridia</i> DTU014 genus <i>Bacteroidetes</i> vadinHA17 genus
	MCL	Bacteroides Petrimonas
	HCL	Caproiciproducens Ruminococcaceae genus Lachnospira Clostridium sensu stricto 1 Methanobacterium Clostridia D8A-2 genus
Thermophilic	control	Defluviitoga Syntrophaceticus Clostridia MBA03 genus Lactobacillus
	HCL	Hydrogenispora Methanosarcina Ruminococcaceae UCG-010 Caproiciproducens Proteiniphilum

Conclusions and Outlook